

## Dengue virus serotype-3 (subtype-III) in Port Blair, India

N. Muruganandam, I.K. Chaaithanya, S. Mullaikodi, P. Surya, R. Rajesh, M. Anwesh, A.N. Shriram & P. Vijayachari

Regional Medical Research Centre (ICMR), Port Blair, Andaman & Nicobar Islands, India

**Key words** *Aedes*; DHF; Port Blair

Epidemics of dengue fever (DF)/dengue hemorrhagic fever (DHF) have been occurring in the South East Asian Region (SEAR)<sup>1</sup> and are considered to be an emerging public health problem world over<sup>2</sup>. Dengue is an *Aedes* transmitted viral infection caused by any of the four dengue virus serotypes (DENV 1–4) which belong to the genus *Flavivirus*. This virus causes spectrum of manifestations ranging from DF to DHF and dengue shock syndrome (DSS), which is fatal and usually associated with secondary infections<sup>1</sup>.

In India, DF, DHF and DSS have been reported from different parts of the country<sup>3–4</sup>. Port Blair is the capital of Andaman and Nicobar Islands, an archipelago of >500 islands and islets situated in the Bay of Bengal, at a distance of 1200 km from the peninsular India. People are constantly moving between mainland of India and these islands. Dengue was not reported from these islands<sup>5</sup>. However, patients with febrile DF, DHF and DSS were reported for the first time in 2009<sup>4</sup>. Subsequently, circulation of dengue serotypes 1 and 2 was established<sup>6</sup>.

From mid-July 2013, medical and public health professionals observed an increase in the number of febrile cases with retro-orbital pain among the Wharf (a structure adjoining the harbour comprising a barrack housing the security personnel) staff of Haddo area adjoining the harbour in Port Blair. The Directorate of Health Services, Andaman and Nicobar Administration requested to investigate the cause for the sudden increase in number of febrile cases. In view of the clinical features suggestive of dengue and widespread prevalence and infestation of vector mosquitoes, dengue infection was suspected<sup>7–8</sup>. We report the findings of the investigations undertaken among the suspected cases (security personnel) residing in barrack adjoining the harbour area.

A case definition for suspected DF was made based on the symptoms/signs listed in WHO guidelines<sup>9</sup>. We used a more inclusive case definition that mandated presence of any one of the symptoms/signs such as sudden onset of fever, headache, retro-orbital pain, and myalgia

for the diagnosis of probable case of dengue fever with an aim to increase the sensitivity.

A total of 23 blood samples were collected from the suspected patients after obtaining informed consent. All the samples were tested for the presence of anti-DENV, anti-chikungunya virus (CHIKV) IgM antibodies by IgM capture ELISA kits developed by National Institute of Virology (NIV), Pune<sup>10</sup>, and anti-leptospiral antibodies by in-house developed microscopic agglutination test (MAT) with a panel of known endemic antigens<sup>11</sup>. All samples were also assessed for IgG antibodies by using IgG capture ELISA (PanBio, France).

RNA was extracted from all the serum samples using Qiagen viral RNA extraction kit (Qiagen, USA) following manufacturer's instructions. RT-PCR analysis was carried out for the presence of DENV and CHIKV RNA

Table 1. IgM, IgG and RT-PCR results of the suspected and confirmed patients

S.No.	Lab. ID	Age/Sex	IgM ELISA	IgG ELISA	RT-PCR
1.	HDW-01	40/M	Positive	Positive	Negative
2.	HDW-02	35/M	Negative	Negative	Negative
3.	HDW-03	55/M	Negative	Negative	Negative
4.	HDW-04	39/M	Negative	Negative	Positive
5.	HDW-05	42/M	Negative	Negative	Positive
6.	HDW-06	36/M	Negative	Negative	Positive
7.	HDW-07	25/M	Negative	Positive	Positive
8.	HDW-08	25/M	Negative	Negative	Positive
9.	HDW-09	24/M	Negative	Negative	Positive
10.	HDW-10	41/M	Negative	Negative	Positive
11.	HDW-11	31/M	Positive	Positive	Negative
12.	HDW-12	35/M	Negative	Negative	Positive
13.	HDW-13	40/M	Negative	Negative	Negative
14.	HDW-14	25/M	Positive	Positive	Negative
15.	HDW-15	42/M	Negative	Positive	Negative
16.	HDW-16	50/M	Negative	Negative	Negative
17.	HDW-17	24/M	Negative	Negative	Negative
18.	HDW-18	25/M	Negative	Negative	Negative
19.	HDW-19	53/M	Negative	Negative	Positive
20.	HDW-20	24/M	Negative	Positive	Negative
21.	HDW-21	44/M	Negative	Negative	Positive
22.	HDW-22	24/M	Negative	Positive	Negative
23.	HDW-23	49/M	Negative	Negative	Negative

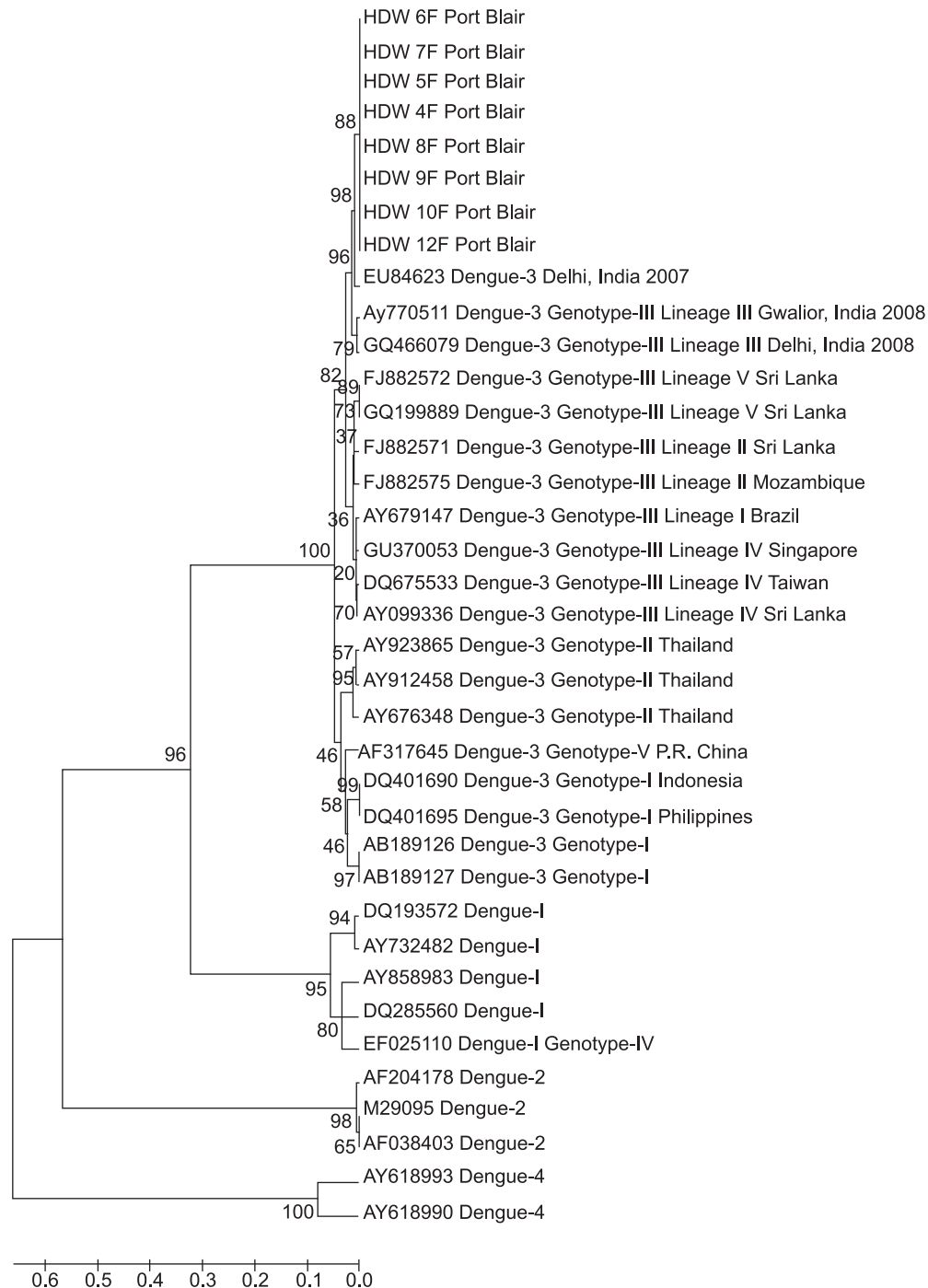


Fig. 1: Phylogenetic neighbour-joining tree showing eight Port Blair DENV sequences (HDW6F, HDW7F, HDW5F, HDW4F, HDW8F, HDW9F, HDW10F and HDW12F) that have been grouped with DENV serotype-3 and genotype-III.

in all the samples by adopting standard protocol<sup>12-13</sup>. Further, PCR product was purified and subjected to DNA sequencing analysis. RT-PCR positive samples were further subjected for virus isolation using Vero-E6 cell lines<sup>14</sup>.

Of the 23 serum samples, 3 (13%) were found positive for DENV specific IgM antibodies. None of the samples were positive for CHIKV IgM and leptospiral

antibodies. Of the 23 samples, 7 (30.4%) were positive for IgG antibodies against dengue virus, 12 (52.1%) samples were positive by RT-PCR analysis, amplified PCR products with the size of 511 bp (Table 1). These findings indicate that the aetiology of the infection was due to DENV. No severe cases of DHF/DSS were observed during the investigations. Patients confirmed to be IgM positives and RT-PCR positive, met the criteria

of dengue fever case definition. The nucleotide sequencing of the 511 bp amplicon confirmed that the virus sequence was homologous with DENV-3. The sequences were submitted to the NCBI (Accession numbers KF915030, KF915031, KF915032, KF915033, KF915034, KF915035, KF915036 and KF915037). All samples were negative for CHIKV RT-PCR analysis. Four RT-PCR positive samples were subjected to culture, for virus isolation but it was not successful.

Further, analyses were carried out by using maximum composite likelihood model<sup>15</sup> and MEGA5<sup>16</sup>. The mean pairwise genetic distance (K2P) between the nucleotide sequences of dengue virus from Port Blair was zero percent. The genetic distance between the dengue virus reported in Delhi (EU846234; 2007) was K2P=0.015%. The mean genetic distance between dengue reported during the current investigation in Port Blair and serotype dengue-3 from worldwide was smaller (0.070%) than that of dengue-1 (0.642%), dengue-2 (1.138%) or dengue-4 (1.356%) serotypes. The neighbour joining phylogenetic tree analysis indicated that the dengue virus associated with the sudden increase in febrile cases clustered with genotype-III and the lineage III (Fig. 1).

Dengue outbreaks with different serotypes have been reported in the mainland, India and also from Andaman Nicobar Islands<sup>3, 6</sup>. Emergence of dengue serotype-3 in north and south India has been reported in the past<sup>17-18</sup>. Earlier studies showed the existence of dengue serotype 1 and 2 in these islands<sup>6</sup>. The investigations from the present study indicate circulation of third serotype of dengue virus in the urban Port Blair. The current findings are in consonance with the investigations reported elsewhere<sup>18</sup>. Thus, it is evident that three serotypes are now in circulation within the urban agglomeration of Port Blair<sup>6</sup>. At this juncture, it is pertinent to mention that as per the antibody-dependent enhancement hypothesis, secondary DENV infections are risk factors for DHF/DSS<sup>19</sup>. Therefore, existence and circulation of multiple serotypes in these islands could precipitate the frequency of occurrences with DSS/DHF. The detection of dengue-3 serotype could be due to introduction of a new serotype; else it could be due to coincidental observation of the circulating serotype. The current scenario warrants constant vigil to monitor the situation coupled with a robust surveillance component integrated with meteorological information and management of *Aedes* spp breeding sites could prevent outbreaks in the future.

#### ACKNOWLEDGEMENTS

The current study formed a part of the activities un-

der the Establishment of Grade I (Diagnostic) Virology at the Regional Medical Research Centre (ICMR), Port Blair which is being supported by extramural grant from the Indian Council of Medical Research (Ref. letter No. 5/8/7/16/2010-ECD-I). The authors are grateful to the Directorate of Health Services, Andaman & Nicobar Administration for extending support during the conduct of study.

#### REFERENCES

1. Pandey BD, Morita K, Khanal SR, Takasaki T, Miyazaki I, Ogawa T, *et al.* Dengue virus, Nepal. *Emerg Infect Dis* 2008; 14: 514-5.
2. Gurugama P, Garg P, Perera J, Wijewickrama A, Seneviratne S. Dengue viral infections. *Indian J Dermatol* 2010; 55: 68-78.
3. Kabilan L, Balasubramanian S, Keshava SM, Thenmozhi V, Sekar G, Tewari SC, *et al.* Dengue disease spectrum among infants in the 2001 dengue epidemic in Chennai, Tamil Nadu, India. *J Clin Microbiol* 2003; 41: 3919-21.
4. Vijayachari P, Singh SS, Sugunan AP, Shriram AN, Manimunda SP, Bharadwaj AP, *et al.* Emergence of dengue in Andaman & Nicobar archipelago: Eco-epidemiological perspective. *Indian J Med Res* 2011; 134: 235-7.
5. Manimunda SP, Singh SS, Sugunan AP, Singh O, Roy S, Shriram AN, *et al.* Chikungunya fever, Andaman and Nicobar Islands, India. *Emerg Infect Dis* 2007; 13: 1259-60.
6. Chaaithanya IK, Bhattacharya D, Muruganandam N, Thamizhmani R, Babu BV, Sundaram SG, *et al.* Dengue: A newly emerging viral infection in Andaman and Nicobar Islands, India. *Epidemiol Infect* 2012; 140: 1920-4.
7. Shriram AN, Sehgal SC. *Aedes aegypti* (L) in Port Blair, Andaman and Nicobar Islands: Distribution and larval ecology. *J Commun Dis* 1999; 31: 185-92.
8. Shriram AN, Sugunan AP, Vijayachari P. Infiltration of *Aedes aegypti* into peri-urban areas in south Andaman. *Indian J Med Res* 2008; 127: 618-20.
9. *Dengue haemorrhagic fever: Diagnosis, treatment, prevention and control*. II edn. Geneva: World Health Organization 1997.
10. Gadkari DA, Shaikh BH. IgM antibodies capture ELISA in the diagnosis of Japanese encephalitis, West Nile and dengue virus infections. *Indian J Med Res* 1984; 80: 613-19.
11. Vijayachari P, Sugunan AP, Sehgal SC. Role of microscopic agglutination test (MAT) as a diagnostic tool during acute stage of leptospirosis in low and high endemic areas. *Indian J Med Res* 2001; 114: 99-106.
12. Lanciotti RS, Calisher CH, Gubler DJ, Chang GJ, Vorndam AV. Rapid detection and typing of dengue viruses from clinical samples by using reverse transcriptase-polymerase chain reaction. *J Clin Microbiol* 1992; 30: 545-51.
13. Hasebe F, Parquet MC, Pandey BD, Mathenge EG, Morita K, Balasubramanian V, *et al.* Combined detection and genotyping of chikungunya virus by a specific reverse transcription-polymerase chain reaction. *J Med Virol* 2002; 67: 370-4.
14. Huhtamo E, Uzcátegui NY, Siikamäki H, Saarinen A, Piiparinen H, Vaheri A, *et al.* Molecular epidemiology of dengue virus strains from Finnish travellers. *Emerg Infect Dis* 2008; 14: 80-3.
15. Tamura K, Nei M, Kumar S. Prospects for inferring very large

- phylogenies by using the neighbour-joining method. *Proc Natl Acad Sci USA* 2004; 101: 11030–5.
16. Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol Biol Evol* 2011; 28: 2731–9.
  17. Dash PK, Parida MM, Saxena P, Abhyankar A, Singh CP, Tewari KN, *et al.* Reemergence of dengue virus type-3 (subtype-III) in India: Implications for increased incidence of DHF and DSS. *Virol J* 2006; 6: 55.
  18. Paramasivan R, Dhananjeyan KJ, Leo SV, Muniaraj M, Thenmozhi V, Rajendran R, *et al.* Dengue fever caused by dengue virus serotype-3 (subtype-III) in a rural area of Madurai district, Tamil Nadu. *Indian J Med Res* 2010; 132: 339–42.
  19. Halstead SB, O'Rourke EJ. Dengue viruses and mononuclear phagocytes. Infection enhancement by non-neutralizing antibody. *J Exp Med* 1977; 146: 201–17.

*Correspondence to:* Dr Paluru Vijayachari, Regional Medical Research Centre (ICMR), Post Bag No.13, Port Blair-744 101, Andaman and Nicobar Islands, India.

E-mail: [directormrc@gmail.com](mailto:directormrc@gmail.com)

*Received:* 30 September 2013

*Accepted in revised form:* 16 January 2014