

Short Research Communication

Viscerotropic potential of parasites isolated from post-kala-azar dermal leishmaniasis cases : An experimental evidence

A.K. Gupta, S. Narayan, N. Verma, A.K. Thakur & P. Das

Rajendra Memorial Research Institute of Medical Sciences (ICMR), Patna, India

Key words India; *Leishmania donovani*; leishmaniasis; viscerotropism

Post-kala-azar dermal leishmaniasis (PKDL) is marked by the appearance of non-ulcerative skin lesions with *Leishmania* parasites, which develop after apparent recovery of kala-azar (KA), though less commonly it has been known to occur in patients who have not suffered previously from visceral leishmaniasis (VL)^{1,2}. This skin reflection of visceral disease (i.e. PKDL) is characterized by hypopigmented macules and erythematous eruptions leading to formation of papules and nodules³ in skin without involvement of viscera. The pathology caused by *Leishmania* parasites in PKDL, is known to be confined to only dermis in the host which raises one of the possibilities for the loss of viscerotropic nature of the parasites. In India, 10% of the patients of kala-azar develop PKDL and now the frequency is declining³. The viscerotropic nature of parasite of PKDL case is poorly understood till date and WHO in its technical report series 949 has recommended that studies are needed to determine the role of PKDL's reservoir for transmission of kala-azar³. The present study was, therefore, undertaken to elucidate this fact and to know whether the parasites which cause dermatotropic lesions still retain viscerotropic or not. The study was approved by the Animal Ethics Committee of the Institute.

Biopsy of dermal lesion (preferably nodular type) of clinically diagnosed PKDL patients was inoculated in modified NNN medium overlaid with Locke's and incubated the culture at $25 \pm 1^\circ\text{C}$ in BOD incubator for one month. Wet smear was examined microscopically on 2–3 days interval for isolation of promastigotes. Newly isolated promastigotes of PKDL cases (n =13, 10 with and 3 without previous history of KA) were adapted by subpassage and proper inoculum size for establishment of infection in susceptible animal.

Promastigotes (1×10^8 cells) of stationary phase of PKDL were inoculated intraperitoneally in inbred Balb/C mice (male, 20–22 g) in a batch (n=5) for each isolate. Animals were sacrificed between 3 and 4 months

to observe the establishment of infection. Dabbed smears of spleen biopsy were stained with leishman stain and examined microscopically for amastigotes. A chopped portion of spleen and liver was inoculated in modified NNN medium overlaid with Locke's medium and wet smear was examined microscopically up to one month at 2–3 days interval for isolation of the promastigotes.

Amastigotes were found in stained dab smears of spleen of all the 13 isolates, as well as promastigotes were also recovered in culture of spleen biopsy of mice of all the isolates. This showed that laboratory inbred Balb/C mice were infected with the PKDL and the PKDL isolates exhibited existence of viscerotropic nature as a change of biological behaviour of parasites in animals. Hence, parasites may not lose their capability to infect viscera and corroborate that the *L. donovani* causing VL and PKDL are the same⁴.

The finding that PKDL cases got VL infection after getting the immunosuppressive infection like measles and repeated attack of malaria and tuberculosis in human⁵ supports our finding of viscerotropic nature of PKDL isolates in animals. In PKDL cases, some scientists have reported the presence of parasites also in bone marrow and lymph node^{6,7} which may indicate the visceralization of parasites. The presence of DNA of *L. donovani* in bone marrow and peripheral blood of PKDL but no evidence of visceral disease⁸ also confirm the transit phase of parasites in between skin and viscera that might be due to immunological hindrance in human. Therefore, our findings confirmed that parasites causing PKDL, retain the biological behaviour of viscerotropism. Thus, in the absence of apparent animal reservoir like in India, PKDL is an important source of parasites to transmit the disease. The work emphasizes the treatment of PKDL to check the transmission of disease⁹. Further studies are warranted to determine the time period of onset of PKDL after treatment of kala-azar cases and how long the parasite remain viable for viscerotropic conditions. There is

also need to ascertain/strengthen the occurrence of cutaneous leishmaniasis by *L. donovani* in Himachal Pradesh (India)¹⁰, Sri Lanka¹¹ and Kenya¹² which may elucidate the link between CL and PKDL.

ACKNOWLEDGEMENTS

Authors are thankful to Mr N.L. Kalra and Dr R.C. Dhiman (NIMR, Delhi) for valuable advice and to Messrs S.K. Chaturvedi, S.K. Sinha and B.N. Roy for technical assistance.

REFERENCES

1. Thakur CP. Review on current status of leishmaniasis: Clinical and laboratory diagnosis. *Proceedings of the Indo-UK Workshop on Leishmaniasis*. New Delhi: Indian Council of Medical Research 1983; p. 50–5.
2. Zijlstra EE, El-Hassan AM, Ismael A. Endemic kala-azar in eastern Sudan: Post-kala-azar dermal leishmaniasis. *Am J Trop Med Hyg* 1995; 52: 299–305.
3. *Control of the leishmaniases: Report of a meeting of the WHO Expert Committee on the control of leishmaniases*. Geneva: World Health Organization 2010. *WHO Tech Rep Ser* 949.
4. El-masum MA, Evans DA. Characterization of *Leishmania* isolated from patients with kala-azar and post kala-azar dermal leishmaniasis in Bangladesh. *Trans R Soc Trop Med Hyg* 1995; 89: 331–2.
5. Nandy A, Addy M, Maji AK, Guha SK, Banerjee D, Chaudhri D. Recurrence of kala-azar after PKDL: Role of co-factors. *Trop Med Int Health* 1998; 3: 76–8.
6. Garg VK, Agarwal S, Rani S, Joshi A, Agarwalla A, Das ML, *et al*. Post kala-azar dermal leishmaniasis in Nepal. *Int J Dermatol* 2001; 40: 179–84.
7. Zijlstra EE, El-Hassan AM. Leishmaniasis in Sudan: Visceral leishmaniasis. *Trans R Soc Trop Med Hyg* 2001; 95 (Suppl 1): S27–58.
8. Meredith SEO, Zijlstra EE, Schoone GJ, Kroon CCM, vans Eys GJJM, Schaeffer KU, *et al*. Development of the polymerase chain reaction for the detection and identification of leishmania parasites, and the application of the PCR for detection of parasites in clinical material. *Arch Inst Pasteur Tunis* 1993; 70: 419–31.
9. Thakur CP, Kumar K. Post-kala-azar dermal leishmaniasis: A neglected aspect of kala-azar control programmes. *Ann Trop Med Parasitol* 1992; 86: 355–9.
10. Sharma NL, Mahajan VK, Kanga A, Sood A, Katoch VM, *et al*. Localized cutaneous leishmaniasis due to *Leishmania donovani* and *Leishmania tropica*: Preliminary. *Trans R Soc Trop Med Hyg* 2005; 72(6): 819–24.
11. Karunaweera ND, Pratloug F, Siriwardane HV, Ihalamulla RL, Dedet JP. Sri Lankan cutaneous leishmaniasis is caused by *Leishmania donovani zymodeme* MON-37. *Trans R Soc Trop Med Hyg* 2003; 97: 380–1.
12. Mebrahtu YB, Eys GV, Guizani I, Lawyer PG, Pamba H, Koech D. Human cutaneous leishmaniasis caused by *Leishmania donovani s.l.* in Kenya. *Trans R Soc Trop Med Hyg* 1993; 87(5): 598–601.

Correspondence to: Dr Shyam Narayan, Microbiology Division, Rajendra Memorial Research Institute of Medical Sciences (ICMR), Agamkuan, Patna–800 007, India.

E-mail: drshyamnarayan@rediffmail.com

Received: 3 July 2012

Accepted in revised form: 14 October 2012