Short Research Communications

Vector abundance and species composition of *Anopheles* mosquito in Calabar, Nigeria

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Malaria is a major cause of morbidity and mortality in Nigeria creating a significant barrier to economic development. Accurate estimation of the extent of the morbidity and mortality is difficult in view of the weakness of the reporting systems for infectious diseases in Africa. In 2008, World Health Organization reported an estimated value of 243 million cases with a mortality of about 863 thousand in the world¹. Malaria transmission in the coastal South States of Nigeria is known to be holoendemic, and perennial. High humidity (80%) and a high mean temperature of 35°C in this area favour the bionomics of the principal malaria vectors (Anopheles gambiae complex and An. funestus)². Vector control is a major component of the Global Malaria Control Strategy (GMCS) and still remains the most generally effective measure to prevent malaria transmission³. However, successful application of vector control measures in a given location requires the understanding of the bionomics of Anopheles species responsible for malaria transmission, including correct and precise identification of the target species⁴ and its distribution⁵.

Studies on malaria entomology in Nigeria are not only few but limited to the north and south-west of the country and are further confounded by the presence of different ecological zones that support breeding of different *Anopheles* species⁶. This study seeks to fill the gaps in the knowledge of bionomics of *Anopheles* from the south zone of the country.

The study was carried out in Calabar Municipality (4°57N, 8°19'E) located in Cross River State – south Nigeria where the vegetation is typically of the rain forest. Mosquitoes were collected using the pyrethrum spray catch (PSC) technique. Collection was carried out twice on the second and fourth week of every month with an average of six houses per area for three localities for 6 months in the morning hours (0600 to 1000 hrs). Between 10 and 15 min after spraying the insecticide, mosquitoes knocked down on white sheets were collected and placed on damp filter papers properly labelled before transfer to the laboratory. Rainfall measurement was carried out by the National Meteorological Centre. Mosquitoes collected were identified with the aid of a stereomicroscope using morphological keys⁷. Each correctly identified *An. gambiae* complex mosquito was kept in dry silica gel and stored at 4°C.

DNA extracted from legs or wings of samples identified as *An. gambiae* were subjected to a species-specific polymerase chain reaction (PCR) assays following the procedure of Scott *et al*⁸. Laboratory strains of the *An. gambiae* complex provided by the Molecular Entomology Laboratory at the Nigerian Institute of Medical Research, Lagos were used as positive controls. PCR products were electrophoresed with 10% ethidium bromide stained agarose gels at 120 volts for 1 h. The amplified fragments were then visualized by illumination with short wave ultra violet trans-illuminator and photographed.

The relationship between the amount of rainfall and vector abundance in the selected area during the experimental period showed a correlation of 0.604. Higher density of mosquitoes was collected between April and June (wet season) than January and March (dry season) as more breeding sites are available in the wet season (data not shown). This has been corroborated by several studies as the rains present favourable environmental conditions that enhance mosquito breeding and survival, through the pro-liferation of larval habitats and improved humidity, respectively⁹.

A total of 675 mosquitoes were caught through PSC. The higher proportion of the culicines (64.1%) compared to the anophelines (35.9%) was thought to be as a result of environmental practices which has subjected most available breeding sites to pollution, hence, encouraging

Study areas	Culicines		Anophelines		Total
	Male	Female	Male	Female	
State housing	46 (25.5)	71 (39.4)	16 (8.8)	47 (26.1)	180
Federal housing	67 (25.5)	92 (35.1)	39 (14.8)	64 (24.4)	262
Old Odukpani	38 (16.3)	123 (52.7)	20 (8.6)	52 (22.3)	233
Total	151 (22.3)	286 (42.3)	75 (11.1)	163 (24.1)	675

Table 1. Morphological identification of mosquitoes caught during the study

Figures in parentheses indicate percentages.

Table 2. Species composition of Anopheles females caught during the study

Study areas	Number caught	Anopheles gambiae complex			An. funestus
		An. gambiae s.s.	An. arabiensis	Unidentified species	complex
State housing	47	34	4	2	7
Federal housing	52	34	6	4	8
Old Odukpani	64	38	8	6	12
Total	163	106 (65)	18 (11)	12 (7.3)	27 (16.7)

Figures in parentheses indicate percentages.

the proliferation of more culicines and reducing the suitability of these sites for anopheline proliferation. The result are presented in Table 1.

Morphological identification of members of the Anopheles collected in this study revealed two major groups with majority (83.4%) belonging to the An. gambiae s.l. and An. funestus complex (16.5%). Molecular characterization of An. gambiae s.l. by polymerase chain reaction assays showed the presence of two sibling species; An. gambiae s.s. (65%) and An. arabiensis (11%) with unidentified species (7.3%) (Table 2). This study confirms the presence of two members of the An. gambiae complex that are capable of transmitting malaria all year round. The predominance of the most efficient vector of malaria An. gambiae s.s. in this study is consistent with available data^{10,11}. An. arabiensis, the least abundance found in this study and other studies^{10,11} has been described as a savannah and dry season zoophilc vectors with a tendency to exhibit exophagic and exophilic behaviour¹².

All the species of *Anopheles* found in this area have been substantially linked to malaria in the country^{11,13}. This study provides a baseline data on the malaria vector species present in Calabar Municipality area giving an insight to the high prevalence of malaria recorded in the region, but more research work is required to properly present and document data on the entomological profile and dynamics of the malaria vectors in this area.

REFERENCES

- 1. *World malaria report* 2009. Geneva: World Health Organization 2009.
- Ezedinach ENU, Ekanem OJ, Chukwuani CM, Meremikwu MM, Ojar EA, Alaribe AAA, Umoton AB, Haller L. Efficacy and tolerability of a low-dose mefloquine-sulfadoxine – pyremethamine combination compared with chloroquine in the treatment of acute malaria infection in a population with multiple drug-resistant *Plasmodium falciparum*. Am J Trop Med Hyg 1999; 6(1): 114–9.
- 3. A global strategy for malaria control. Geneva: World Health Organization 1993; p. 1–30
- Coluzzi M. Malaria and the Afro-tropical ecosystem; effect of a man-made environmental changes. *Parasitologia* 1994; 35: 223–7.
- Coetzee M, Craig M, leSueur D. Distribution of African malaria mosquitoes belonging to the *Anopheles gambiae* complex. *Parasitol Today* 2000; *16*(2): 74–7.
- 6. Awolola TS, Bitsindou P, Bagoyoko M, Manga L. *Malaria Entomological Profile for Nigeria* 2008.
- Gillies MT, deMeillon B. *The Anophelinae of Africa south of the Sahara*, II edn. Johannesburg, South Africa: Publications of the South African Institute for Medical Research, No. 54, 1968; p. 7–13.
- Scott JA, Broodon WG, Collins FH. Identification of single specimens of the *Anopheles gambiae* complex by the polymerase chain reaction. *Am J Trop Med Hyg* 1993; *49*(4): 520–9.
- Minakawa N, Mutero CM, Githure JI, Beier JC, Yan G. Spatial distribution and habitat characterization of anopheline mosquito larvae in Western Kenya. *Am J Trop Med Hyg* 1999; *61:* 1010–6.

- Alaribe AAA, Ejezie GC, Ezedinachi ENU. Molecular identification of *Anopheles gambiae* complex found in Calabar, using the polymerase chain reaction technique. *Mary Slessor J Med* 2003; 3(1): 61–4.
- Okwa OO, Rasheed A, Adeyemi A, Omoyeni M, Oni L, Fayemi A, Ogunwomoju A. *Anopheles* species abundances, composition and vectoral competence in six areas of Lagos, Nigeria. *J Cell Animal Biol* 2007; 1(2): 19–23.
- 12. Randrainasolo DO, Coluzzi M. Genetic investigations in zoophilic and endophilic *An. arabiensis* from Antonanvo area (Madagascar). *Parasitologia* 1989; 29: 93–7.
- Awolola TS, Ibrahim K, Okorie T, Koekemoer LL, Hunt RH, Coetzee M. Species composition and biting activities of anthropophilic *Anopheles* mosquitoes and their role in malaria transmission in a holoendemic area of south-western Nigeria. *Afr Entomol* 2003; *11*(2): 227–32.

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