Short Research Communications

Variations in species composition and infection rates in *Anopheles* gambiae s.l. across eco-vegetational zones in Nigeria and Cameroon

M.A.E. Noutcha^{a,b} & C.I. Anumudu^a

^aCellular Parasitology Programme (CPP), Department of Zoology, University of Ibadan, Ibadan, Oyo State; ^bDepartment of Animal and Environmental Biology, University of Port Harcourt, Port Harcourt, Rivers State, Nigeria

Key words Anopheles gambiae s.l.; eco-vegetational zone; infection rates; species composition

Significant variations in vector biology within and between countries have been reported in malaria epidemiological studies¹. It is now generally agreed that a clear understanding of the detailed epidemiology of the disease is a pre-requisite to effective malaria control in the African sub-region². Anopheles gambiae complex (Diptera: Culicidae) is one of the most efficient vectors of Plasmodium falciparum in the African sub-region; it is a complex of sibling taxa³. Two of these sibling species, An. gambiae sensu stricto and An. arabiensis are the major vectors⁴. Relative abundance of the sibling species varies across eco-vegetational zones^{5–8}. Vector competence among sibling species is dependent on the degree of anthropophily, endophily and ability to recover after the dry season⁵. Consequently, variations occur in infection rates among sibling species 9-10. Two of the problems of malaria vector control in Nigeria are the diversity of anopheline vectors and large size of the country. Infection rates varied across ecological zones in Nigeria¹¹ and Cameroon¹². This report focuses on species composition and infection rates from investigations conducted to obtain key entomological indices of An. gambiae s.l. in four villages across eco-vegetational zones in Nigeria (Igbo-Ora and Bama) and Cameroon (Fokoué and Mora). Malaria is perennial and endemic at Igbo-Ora, Bama, Mora and Fokoué, with transmission reaching its peak in August; the density and activity of An. gambiae s.l. in these villages follow the rainfall patterns ¹³.

Bama (13.68° E, 11.16° N) and Mora (14.68° E, 11.05° N), villages in the Lake Chad Basin are in the Sudan savanna; Igbo-Ora (3.28° E, 7.43° N) in the Savanna woodland ecotone, between the forested south and the Savanna, while Fokoué (10.12° E, 5.35° N) is located in the hilly savanna woodland of central Cameroon. There are two overlapping seasons in the forest and savanna woodland: November–April (dry), May–October (rainy); the Sudan savanna is characterized by three seasons: hot-dry (February–May); rainy (June–September) and dry humid (October–January). Detailed description of Igbo-Ora has been documented elsewhere 14.

The four villages were surveyed and marked into quarters containing groups of houses. The rivers and streams that traverse the villages served as key landmarks for the selection of collection sites. Mosquitoes were collected weekly from May 2004-October 2005 to cover the dry and rainy seasons. Mosquitoes were caught using an aspirator from residual fauna of indoor resting mosquitoes during the periods: 0800-1100 hrs and 1600-1900 hrs. Sample processing, analyses and data acquisition for the identification of sibling species and determination of infection rates were done. Sibling species were identified using PCR¹⁵ and infection obtained by the circumsporozoite (CSP) antigen by sandwich ELISA¹⁶. The Chi-square and Student's t-test were used for statistical analyses.

Table 1. Numbers of mosquitoes collected from study villages of Nigeria and Cameroon

Village	Species	No. collected (%)			
		2004	2005		
Bama	An. arabiensis	25 (58.1)	136 (100)		
	An. gambiae s.s.	18 (41.9)	0 (0)		
Igbo-Ora	An. arabiensis	216 (55.4)	68 (49.3)		
	An. gambiae s.s.	174 (44.6)	70 (50.7)		
Mora	An. arabiensis	1 (100)	11 (91.7)		
	An. gambiae s.s.	0 (0)	1 (8.3)		
Fokoué	An. arabiensis	0 (0)	0 (0)		
	An. gambiae s.s.	0 (0)	5 (100)		

In the arid Sudan savanna, Bama (Nigeria) and Mora (Cameroon), An. arabiensis was more abundant than An. gambiae s.s., constituting 58.1–100% (Table 1). When the data from both villages were used, the difference in abundance was significant in both years: 2004 ($\chi^2 = 0.708$; p < 0.05) and 2005 ($\chi^2 = 11.41$; p < 0.05). Pooling of the 2004 and 2005 data for each village showed that the differences were also significant: Bama ($\chi^2 = 63.30$; p < 0.05) and Mora ($\chi^2 =$ 0.07; p < 0.05). In the savanna-woodland, Igbo-Ora (Nigeria) and Fokoué (Cameroon), annual differences in species abundance of the pooled data from both villages were significant: 2004 ($\chi^2 = 390$; p < 0.05), $2005 (\chi^2 = 4.97; p < 0.05)$. When the 2004 and 2005 data were pooled in each village, the differences were also significant: Igbo-Ora ($\chi^2 = 1.53$; p < 0.05) and Fokoué (t = 1; p < 0.05).

At Bama, in the arid Sudan savanna, the infection rate in An. arabiensis was higher in 2004, but the difference was not significant (t = 1; p > 0.05). In 2005, no infective mosquito was recorded in both species (Table 2). At Igbo-Ora in the savanna woodland, the infection rate in An. arabiensis was higher in 2004, but in 2005, the infection rate in An. gambiae s.s. was higher. However, the pooled 2004 and 2005 data did not reveal any significant difference in infection rates between species ($\chi^2 = 0.48$; p > 0.05). The significantly higher number of An. arabiensis in the Sudan savannah of Cameroon and Nigeria is in agreement with the conclusions of earlier studies that An. arabiensis is more adapted to drier habitats 12,17 . However, the higher number of An. arabiensis at Igbo-Ora, approximately 70 km from coastal Lagos¹⁸ was evidence of an advance of the species to more humid areas in West Africa. This southward advance of An. arabiensis had also been observed in Cameroon where Domfang et al^{19} recorded An. gambiae s.s., An. arabiensis and An. funestus at Tibati, a village in the savanna woodland zone. The observed southward advance of An. arabiensis highlights the limitation in the exclusive reliance on climate and distributional data or the use of spatial imaging to predict species occurrence^{6,17}.

The higher infection rate of *An. arabiensis* in the Sudan savanna was not surprising because it had been reported as contributing a large proportion of the infectious mosquito bites on the African continent^{10,20–21}. The northward advance of *An. gambiae*

Table 2. Infection rates of *Anopheles gambiae s.s.* and *An. arabiensis* at villages in the Sudan savanna and savanna woodland, Nigeria

Village	Species	Periods						
		2004			2005			
		No. examined	No. positive	Infection rate	No. examined	No. positive	e Infection rate	
Bama	An. arabiensis	25	1	4	0	0	0	
	An. gambiae s.s.	18	0	0	0	0	0	
Igbo-Ora	An. arabiensis	216	5	2.31	68	1	1.47	
	An. gambiae s.s.	174	2	1.15	70	2	2.85	

s.s. into the arid north at Bama in 2004, reflected the plasticity of the species, which was highlighted by White⁵. The extremely low mosquito numbers at Mora might have been due to local behavioural peculiarities, such as exophily after a blood meal. The low numbers recorded at Fokoué was probably associated with low temperatures in the hilly terrain. The non-significant difference in the infection rates recorded in An. gambiae s.s. and An. arabiensis from the combined data of 2004 and 2005, indicates the importance of both species of the An. gambiae complex in malaria transmission in the arid north or humid south of West Africa, although An. gambiae s.s. had often been considered a more competent vector⁴.

References

- 1. Collins FH, Paskewitz SM. Malaria: current and future prospects for control. *Annual Rev Entomol* 1995; *40*: 195–219.
- 2. Touré YT, Petrarca V, Traoré SF, Coulibaly A, Maiga HM, Sankare O, Sow M, Coluzzi M. The distribution and inversion polymorphism of chromosomally recognized taxa of the *Anopheles gambiae* complex in Mali, West Africa. *Parassitologia* 1998; *40:* 477–511.
- 3. Coluzzi M. Heterogeneities of the vectorial malaria system in tropical Africa and their significance in malaria epidemiology and control. *Bull WHO* (Suppl) 1984; 62: 483–97.
- 4. Constantini C, Gibson G, Sagnon N. Mosquito responses to carbon dioxide in a West African Sudan savanna village. *Med Vet Entomol* 1996; *10*(3): 220–7.
- 5. White GB. *Anopheles gambiae* complex and disease transmission in Africa. *Trans R Soc Trop Med Hyg* 1974; 68: 278–301.
- 6. Lindsay SW, Parson I, Thomas CJ. Mapping the ranges and relative abundance of the two principal African malaria vectors, *Anopheles gambiae* sensu stricto and *Anopheles arabiensis* using climatic data. *Proc R Soc B. Biol Sci* 1998; 265: 847–54.
- 7. Coetzee M, Craig M, Le Sueur D. Distribution of African malaria mosquitoes belonging to the *Anopheles gambiae* Complex. *Parasitol Today* 2000; *16*: 74–7.
- 8. Coetzee M. Distribution of the African malaria vectors of the *Anopheles gambiae* complex. *Am J Trop Med Hyg* 2004; 70 (2): 103–4.

- 9. Taylor KA, Koros JK, Nduati J, Copeland RS, Collins FH, Brandling-Bennett D. *Plasmodium falciparum* infection rates in *Anopheles gambiae*, *An. arabiensis* and *An. funestus* in western Kenya. *Am J Trop Med Hyg* 1990; *43* (2): 124–9.
- 10. Fontenille D, Lochouarn L, Diatta M, Sokhna C, Dial I, Diagne N, Lemasson JJ, Ba K, Tall A, Rogier C, Trape JF. Four years: entomological study of the transmission of seasonal malaria in Senegal and the bionomics of *Anopheles gambiae* and *Anopheles arabiensis*. *Trans R Soc Trop Med Hyg* 1997; 91 (3): 270–2.
- 11. Okwa OO, Akinmolaran FJ, Carter V, Hurd H. Transmission dynamics of malaria in four selected ecological zones of Nigeria in the rainy season. *Ann Afr Med* 2009; 8(1): 1–9
- Wondji CS, Simard F, Petrarca V, Etang J, Santalomazza F, della Torre A, Fontenille D. Species and population of the *Anopheles gambiae* complex in Cameroon with special emphasis on chromosomal and molecular forms of *Anopheles gambiae* s.s. *J Med Entomol* 2005; 42(6): 998–1005.
- Touré M. Thèse de Médécine M100 bibliothèque Faculté de Médécine Pharmacie et Odonto-Stomatologie, Université du Mali. 2002.
- 14. Noutcha MAE, Anumudu C. Entomological indices of *Anopheles gambiae* sensu lato at a rural community in southwest Nigeria. *J Vector Borne Dis* 2009; 46 (1): 43–51.
- 15. Scott JA, Brongdon WG, Collins FH. Identification of single specimen of the *Anopheles gambiae* complex by the polymeraze chain reaction. *Am J Trop Med Hyg* 1993; 49: 520–9
- 16. Wirtz RA, Zavala F, Charoenvit Y, Campbell GL, Burkott TR, Schneider I, Esser KM, Beaudoin RL, Andre RG. Comparative testing of monoclonal antibodies against *P. falciparum* sporozoïtes for ELISA development. *Bull de l' OMS* 1987; 65: 39–45.
- 17. Levine RS, Peterson AT, Benedict MQ. Geographic and ecologic distributions of the *Anopheles gambiae* complex predicted using a genetic algorithm. *Am J Trop Med Hyg* 2004; 70(2): 105–9.
- 18. Encarta. *The World Atlas* 2005 (Available from: *http://www.encarta.com*).
- Domfang J, Wondji C, Awono A, Antonio-Nkodjo, Tato J, Simard F. Malaria transmission dynamics and the diversity of anopheline vector populations in Tibati, Cameroon. *Acta Trop* 2005; 95S: S1–S506.
- 20. Diatta M, Spiegel A, Lochouarn L, Fontenille D. Similar feeding preferences of *Anopheles gambiae* and *A*.

- arabiensis in Senegal. Trans R Soc Trop Med Hyg 1998; 92: 270–2.
- 21. Kent RJ, Thuma PE, Mharakhurwa S, Norris DE. Seasonality, blood feeding behavior and transmission of *Plasmodium falciparum* by *Anopheles arabiensis* after an extended drought in southern Zambia. *Am J Trop Med Hyg*

2007; 76 (2): 267–74.

22. Ndiah MO, Berengues C, Konate L, Sokhna C, Boudin C, Trape JF, Fontenille D. Dynamics of transmission of *Plasmodium falciparum* by *Anopheles arabiensis* and the molecular forms of *Anopheles gambiae* in Dielmo, Senegal. *Malar J* 2008; 7: 136–42.

Corresponding author: M.A.E. Noutcha, Department of Animal and Environmental Biology, University of Port Harcourt,

P.M.B. 5323, Choba, Port Harcourt, Rivers State, Nigeria.

E-mail: naemekeu@yahoo.com

Received: 20 August 2009 Accepted in revised form: 26 January 2010